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Simultaneously enhanced biopolymers production and sludge dewaterability of waste activated sludge by synergetic integration process of short-time aerobic digestion with cocoamidopropyl betaine and calcium oxide

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Graphical Abstract



1	Simultaneously enhanced biopolymers production and sludge dewaterability of
2	waste activated sludge by synergetic integration process of short-time aerobic
3	digestion with cocoamidopropyl betaine and calcium oxide
4	
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16 Abstract

17 Waste activated sludge (WAS) has seriously threatened the environment safety 18 and the public health due to its rapid growth and complex components. 19 Simultaneously enhanced the biopolymers production and the sludge dewaterability of 20 WAS were investigated by synergetic integration process of the short-time aerobic 21 digestion (STAD) with cocoamidopropyl betaine (CAPB) and calcium oxide (CaO). 22 STAD could improve the biopolymers production by biosynthesis. CAPB could 23 further significantly enhance the biopolymers production and optimized the CaO (0.1~0.2 g/g TSS) could dramatically enhance the sludge 24 constituents. 25 dewaterability by forming a multi-grid skeleton in WAS, while the biopolymers 26 production could almost remain stable. Especially, the synergetic integration process 27 of STAD with 0.1 g CAPB/g TSS for 8 h and 0.1 g CaO/g TSS could cost-effectively 28 enhance both the biopolymers production and the sludge dewaterability. The 29 produced biopolymers showed strong adsorbability for heavy metals (eg, 375 mg Accordingly, the developed novel process is of big Cu^{2+}/g biopolymers). 30 significance for resource utilization and volume reduction of WAS. 31

32

33 Keywords: Waste activated sludge (WAS); Short-time aerobic digestion (STAD);
34 Sludge dewaterability; Cocoamidopropyl betaine (CAPB); Calcium oxide (CaO)

35 **1 Introduction**

Waste activated sludge as a typical by-product is largely produced in wastewater 36 treatment plants (WWTPs) (Kiff, 1978). According to the report of development 37 prospect and investment forecast on China sewage treatment industry in 2017-2021 38 39 ((EIC), 2017), the yield of waste activated sludge (WAS) with 80% water content in urban WWTPs reached 40.88 million tons in 2015, which will further dramatically 40 increase due to the advancement of the urbanization level. WAS has been seriously 41 42 threatened the environment and public health, and it's imperative to develop effective 43 processes for resource utilization and volume reduction of WAS (Campbell, 2000; 44 Paul et al., 2006).

WAS contains about 60% of organic matters including proteins (PN), 45 46 polysaccharides (PS), nucleic acids (NA), humic acids (HA) etc., and another 40% of inorganic matters contain calcium and magnesium salt (Fytili and Zabaniotou, 2008). 47 Because of these organic and inorganic matters, WAS could be used as the source 48 49 materials of fertilizer, building material, activated carbon, biopolymers for heavy 50 metals removal and so on (Tyagi and Lo, 2013). Although aerobic digestion is widely utilized in middle and small WWTPs, the biomass is majorly mineralized by 51 52 consuming lots of energy (Zhang et al., 2016). Short-time aerobic digestion (STAD) was found to improve the biopolymers production from WAS by biosynthesis. The 53 54 biopolymers are excellent biosorbents for removing and recycling heavy metals from wastewater due to their large amounts of negatively charged functional groups such as 55 56 carboxyl, hydroxyl, amino, phosphate and sulfate (More et al., 2012; Zhou et al., 2015; 57 Zhou et al., 2016a). Surfactants like cocoamidopropyl betaine (CAPB), were further

found to promote the STAD of WAS to enhance the biopolymers production via
forming micelles effective for improving the solubility of the macromolecular organic
matters in EPS and IPS (Sengco et al., 2001; Xia et al., 2017; Zhou et al., 2017d).
However, the moisture in the digested sludge after extracting the biopolymers was
still high (Neyens et al., 2004).

The moisture in WAS contains free moisture, interstitial moisture, surface 63 moisture and bound moisture (Tsang and Vesilind, 1990). 64 Bound moisture is chemically bound to the solid phases in WAS, which contain extracellular polymeric 65 66 substances (EPS) and intracellular polymeric substances (IPS) (Vaxelaire and Cézac, 67 2004). Unlike other three kinds of moistures, bound moisture proportion is normally lower than 3% in the total moisture of WAS, but which cannot been removed using 68 69 the mechanical force separation methods containing gravitational settlement, 70 centrifugation and negative pressure separation (Tsang and Vesilind, 1990; Lee, 1996). In order to convert the bound moisture stored in EPS and IPS to free moisture and 71 achieve the fast solid and liquid separation, solubilization of EPS and even cell lysis 72 that resulted to the release of IPS should be the significant step. Previous studies 73 (Neyens et al., 2002; Neyens and Baeyens, 2003a, b; Brar et al., 2010) mentioned that 74 75 thermal hydrolysis (acid, neutral and alkaline conditions) and chemical oxidation 76 (advanced oxidation, ozonation, H₂O₂ oxidation, nanomaterials catalytic oxidation 77 and so on) could be used to solubilize EPS and even cause cell lysis effectively. Additionally, as the commonly used multi-grid skeleton materials, calcium oxide 78 (CaO), flyash and diatomite could be used to enhance the strength and reduce the 79 80 compressibility of WAS, and then further enhance the dewaterability of WAS (Shi et

al., 2008; Thapa et al., 2009). In particular, CaO as a cheap dehydrant can react with
water in WAS and raise the sludge temperature, cause cell lysis and kill pathogenic
bacteria at the same time, which result in better dewaterability and stabilization of
WAS (Zhai et al., 2012; Liu et al., 2015). Thus, CaO might be an optimal choice to
further decrease the moisture in the digested sludge after extracting the biopolymers.

In this study, simultaneously enhanced biopolymers production and sludge 86 dewaterability of WAS were investigated by comparing the synergetic integration 87 process of STAD with CAPB and CaO with other two processes (CAPB+STAD, 88 STAD+CaO). The effects of the three processes on the normalized CST of WAS 89 and water content of sludge cake, and the production of PN, PS and NA in the 90 biopolymers. The biopolymers with various proportions of fractions were used to 91 test their absorbability for Cu^{2+} and the key fraction of the biopolymers responsible 92 for binding Cu^{2+} were identified. Scanning electron microscopy (SEM) was used to 93 94 characterize the variation of WAS surface structure after processed by the three 95 methods. Mechanisms of simultaneously enhancing biopolymer productions and 96 sludge dewaterability by the synergetic process of CAPB+STAD+CaO were 97 elaborated.

98 2 Materials and Methods

99 2.1 Chemicals and waste activated sludge samples

100 Cocoamidopropyl betaine surfactant ($C_{19}H_{38}N_2O_3$, CAPB), with a molecular 101 weight of 342.52 g/mol, was purchased from Shanghai Chem. Co. Ltd., China. 102 Cu(NO₃)₂·3H₂O (>99% purity) was obtained from Runjie Chemistry Reagent 103 (Shanghai, China) and the stock solution of Cu²⁺ was prepared by dissolving 104 Cu(NO₃)₂·3H₂O in distilled water at an initial concentration of 1000 mg/L.

WAS samples were obtained from the secondary sludge settling tank in a full-scale municipal wastewater treatment plant in Shanghai, China. Collected samples were screened through a 1.2 mm sieve to remove grit and then gravity concentration for 30 min. Supernatants were removed and the main parameters of concentrated sludge were shown in Table 1. Concentrated sludge was stored at 4 °C and used within 2 days.

111 2.2 Three processes of WAS

For the CAPB+STAD process experiments, CAPB with the dosage of 0.00, 0.02, 0.05, 0.10, 0.15 and 0.20 g/g TSS were added to the concentrated WAS and then aerobic digested in six plastic cylindrical reactors simultaneously. The size parameters and schematic configuration of the aerobic digestion reactor, and the aerobic digestion of WAS were described in detail in our previous studies (Zhang et al., 2016; Zhou et al., 2017b).

For the STAD+CaO process experiments, WAS without adding CAPB was initially aerobic digested for 8 hours and CaO was added to the digested sludge with the dosage of 0.0, 0.1, 0.2, 0.3, 0.4 and 0.5 g/g TSS, and then mixing the samples at

121	250 rpm for 2 min, followed by slow stirring at 150 rpm for 8 min with the
122	temperature of 298.15 \pm 1 K using a constant temperature – magnetic stirrer (RCT/
123	RET, Guangzhou Boletai. Co., China).
124	For the synergetic integration process (CAPB+STAD+CaO) experiments, WAS
125	with CAPB (0.10 g/g TSS) was initially aerobic digested for 8 hours, and then CaO
126	was added to the digested sludge with the dosage of 0.0, 0.1, 0.2, 0.3, 0.4 and 0.5 g/g
127	TSS. The samples were mixed immediately using the same method as presented
128	above.
129	2.3 Biopolymer extraction and sludge dewatering
130	The vacuum filtration method was used for simultaneously biopolymer
131	production and sludge dewatering. 100 mL of processed sludge was poured into a
132	Buchner funnel (Figure S1), fitted with pre-wetted quantitative filter paper (Cat No.
133	92410552S), under a constant vacuum pressure of 0.045 MPa supplied by a vacuum
134	pump (DAP-12, Ulvac Kiko Inc., Japan) for 30 min or until the vacuum could not be
135	maintained (< 30 min). The sludge cake on the surface of the filter paper was dried
136	at 105 °C until obtained a constant weight. The collected filtrates were centrifuged
137	twice at 10800×g with a constant temperature of 277.15 K for 10 min. The
138	supernatant was the raw biopolymer, which was further purified via cold acetone
139	method (Zhou et al., 2015).

140 2.4 Adsorption of Cu^{2+} by the biopolymer

The biopolymers used in the adsorption experiments contain three parts: from the
CAPB+STAD process with the dosage of CAPB 0.10 g/g TSS and aerobic digestion
time 2, 4, 8, 12 and 24h (Part I); from the STAD+CaO process experiments with the

dosage of CaO 0.1, 0.2, 0.3, 0.4 and 0.5 g/g TSS (Part II); and from the synergetic 144 integration process (CAPB+STAD+CaO) experiments with the dosage of CAPB and 145 aerobic digestion time 0.10 g/g TSS and 8 h, respectively, and the dosage of CaO 0.3 146 Adsorption experiments were carried out by adding the g/g TSS (Part III). 147 biopolymers into 250-mL Erlenmeyer flasks containing 50 mL of Cu(NO₃)₂ solutions 148 with the initial concentration of 10 mg Cu^{2+}/L and weight ratio of biopolymer/ Cu^{2+} at 149 The flasks were sealed immediately after adding the biopolymers to decrease 150 2.0. Reaction conditions were controlled as follows: 151 the evaporation of solution. temperature 303.15 K, initial pH 5.5, isothermal shaking speed 150 rpm, and reaction 152 Our previous studies (Zhang et al., 2014; Zhou et al., 2016c) 153 time 30 min. demonstrated that the adsorption of Cu^{2+} by the biopolymers almost reached 154 equilibrium after 30 min. Samples were taken from the solutions after agitation and 155 separated from the precipitates prior to analysis through centrifugation at $10,800 \times g$ 156 for 10 min with a temperature of 277.15 K, and then filtrated using 0.22-µm cellulose 157 nitrate membrane filters. Filtrate was acidified with HNO₃ and then analyzed the 158 residual Cu²⁺ using inductively coupled plasma atomic emission spectrometry 159 (ICP-AES, Agilent 720ES, USA). The adsorbed Cu^{2+} by the biopolymers (q_e , mg/g) 160 is calculated using Eq.(1) (Zhou et al., 2016b; Zhou et al., 2017c). 161

$$q_e = \frac{(C_0 - C_e)V}{W} \tag{1}$$

where C_0 and C_e are the initial and equilibrium concentration of Cu^{2+} in the solution, respectively, mg/L; V is the volume of the solution, L; and W is the weight of the biopolymers, g. 165 *2.5 Analytical methods*

The pH of sludge samples were measured with a pH meter (HQ40d, Hach, USA). 166 Water content of sludge cake, TSS and VSS of WAS were analyzed using the 167 standard methods (American Public Health et al., 1915). The dewaterability of 168 sludge was determined by measuring the capillary suction time (CST) with a CST 169 instrument and a Whatman No. 17 chromatography grade paper, and then divided by 170 the initial TSS for normalization (APHA, 2005). SEM image of the sludge samples 171 were completed using Philips XL 30 ESEM. PN, PS and NA in the biopolymers 172 were examined using the following colorimetric methods: Bradford method for PN 173 content, bovine serum albumin as the standard (Frølund et al., 1996); phenol-sulfuric 174 acid method for PS content, glucose as the standard (Frølund et al., 1996); 175 diphenylamine colorimetric method for NA content, calf thymus deoxyribonucleic 176 acid as the standard (Frølund et al., 1996). 177

178 2.6 Statistical analysis

Triplicate processes of activated sludge and the adsorption experiments of the 179 biopolymers for Cu^{2+} were carried out at each condition, and each sample from each 180 experiment was assayed one time for pH, TSS, VSS, CTS, water content of sludge 181 cake, PN, PS, NA and residual Cu²⁺. Results are expressed as the mean and standard 182 deviation of the three measured samples (mean \pm SD). Pearson's correlation 183 coefficient was used to evaluate the linear correlation between the dosage of CAPB 184 and the fractions concentration in the biopolymers under various aerobic digestion 185 times, and the adsorption capacity of Cu²⁺ and the proportions of PN, PS and NA in 186

- the biopolymers. Correlations were considered statistically significance with a 95%
- 188 confidence interval (P < 0.05).

189 3 Results and discussion

190 3.1 Effects of CAPB on biopolymers production and sludge dewaterability of WAS by
191 STAD

192 Fig.1 (a)-(d) shows the effects of CAPB dosage on the production of PN, PS, NA and total biopolymers, respectively, and Fig.S2 shows the corresponded proportions 193 194 of PN, PS, NA and biopolymers at the noted STAD times in the CAPB+STAD process of WAS. Table 2 presents the parameters of the linear relationship between 195 196 CAPB dosage and the biopolymers concentrations and its fractions. CAPB could 197 enhance the solubilization of organic matters from WAS and resulted in the dramatic increase of the biopolymers, which also gradually increased with the increasing of 198 CAPB dosage. CAPB dosage also had a strong and nearly linear relationship with 199 200 the biopolymers and its fractions concentrations as all of the correlation coefficients (R^2) were above 0.95 (Table 2). PN were the key fraction and it also keep increasing 201 as CAPB dosage increased, and PS was higher than NA as CAPB dosage lower than 202 203 10% (Fig.S2). While, NA was higher than PS as CAPB dosage higher than 10%, 204 meaning cell lysis occur. During the STAD process, K, meaning the release of organic matters, gradually increased in the initial 8 h, but gradually decreased in the 205 later stage (Table 2), which was consistent with the biopolymers and its fractions 206 variations (Fig. 1). The long alkyl chain in CAPB can form micelles that could 207 enhance the solubility of macromolecular organic matters (MOM) in WAS (Zhou et 208 al., 2017a; Zhou et al., 2017d) and resulted in the biopolymers increase in the initial 8 209 210 h. Moreover, soluble biopolymers could be consumed by heterotrophic bacteria as

which could be the proper carbon sources and electron donor (Zevin et al., 2015),

which was responsible for the biopolymers decrease in the later stage of STAD.

Fig.1(e)-(f) shows the effects of CAPB dosage on the normalized CST of WAS 213 and water content of sludge cake at the noted STAD times. Fig.2 presents the 214 Pearson correlations between the normalized CST and the PN, PS, NA and 215 biopolymers concentrations. The normalized CST of WAS gradually increased with 216 the increasing of CAPB dosage, meaning a deteriorated dewatering performance of 217 WAS after adding CAPB. Especially, CAPB dosage higher than 10% caused a 218 dramatically increase of the normalized CST, which also keep increasing during the 219 STAD process. The normalized CST had a strong linear positive relationship with the 220 concentrations of PN ($R^2 = 0.854$, P < 0.01), PS ($R^2 = 0.788$, P < 0.01), NA ($R^2 =$ 221 0.768, P < 0.01) and biopolymers ($R^2 = 0.819$, P < 0.01). CAPB could enhance the 222 solubilization of MOM containing EPS and even IPS in WAS, and high concentration 223 of soluble organic matters in the mixed culture caused the deteriorated dewatering 224 performance of WAS. Previous studies (Chen et al., 2001; Zhou et al., 2017d) also 225 226 mentioned that hydrophilic organic matters could adsorb water and high concentration of soluble organic matters in the mixed culture led to the poor dewatering 227 performance of WAS. CAPB could significantly increase the water content of 228 sludge cake, lead to the deteriorated dewatering content of WAS. During the STAD 229 process, the water content of sludge cake gradually decreased in the initial stage, but 230 which increased again later. Results should that STAD could enhance the 231 dewaterability of WAS, which is consistent with previous findings (Zhang et al., 232 2016). 233

234 *3.2 Effects of CaO on biopolymers production and sludge dewaterability of WAS by*

235 *STAD*

Fig.3 (a)-(b) shows the effects of CaO dosage on the biopolymers and its 236 fractions production and the variation of fractions proportion in the STAD+CaO 237 process of WAS, respectively. PN, PS, NA and the biopolymers dramatically 238 increased as CaO dosage increased from 0 to 0.3 g/g TSS, but which gradually 239 decreased for a higher CaO dosage. The maximum biopolymers and proteins 240 concentrations were 10.5 and 25.59 mg/gVSS, respectively. PN (41%~52%) were 241 the key fraction in the biopolymers, followed by PS (35%~41%) and NA (13%~23%). 242 Fig.3 (c)-(d) presents the normalized CST of WAS and water content of sludge cake. 243 244 Normalized CST slightly increased as CaO dosage increased from 0 to 0.3 g/g TSS, 245 but which dramatically decreased for a higher CaO dosage. Water content of sludge cake dramatically decreased from 78.1% to 73.5% after adding 0.1 g/g TSS of CaO, 246 and which further gradually decreased after adding more CaO. Compared with 247 CAPB+STAD process, adding CaO could significantly reduce the water content of 248 sludge cake, but the biopolymers concentrations dramatically decreased. 249

250 *3.3 Synergetic effects of CAPB and CaO on the biopolymers production and WAS*

251 *dewaterability of WAS by STAD*

Fig.4 (a)-(b) shows the synergetic effects of CAPB and CaO on the biopolymers production and its fractions, and the fractions proportion in the biopolymers of the synergetic integration process (CAPB+STAD+CaO) of WAS, respectively. Sludge was treated by aerobic digested for 8h with 0.10 g CAPB /g TSS, and then treated by various dosages of CaO. Concentrations of the biopolymers and its fractions almost

257 remained stable as CaO dosage increased from 0 to 0.2 g/g TSS, but which gradually decreased as CaO dosage higher than 0.2 g/g TSS. Proportions of the fractions 258 almost remained stable and PN (61%~64%) were the key fraction in the biopolymers, 259 260 followed by PS (24%~26%) and NA (11%~15%). Fig.4 (c)-(d) presents the normalized CST of WAS and water content of sludge cake after processed by CAPB 261 and CaO. Normalized CST slightly increased as CaO dosage increased from 0 to 0.1 262 g/g TSS, but which dramatically decreased for a higher CaO dosage. Water content 263 of sludge cake dramatically decreased from 78.3% to 74.9% after adding 0.1 g/g TSS 264 of CaO, and which also further gradually decreased after adding more CaO. 265

Results should that low dosage of CaO in CAPB+STAD+CaO process has 266 minimal effects on the biopolymers concentration, which was also similar with that 267 after CAPB+STAD process but much higher than that by STAD+CaO process, and 268 PN proportion is much higher than the separate process by CAPB+STAD or 269 STAD+CaO. Water content of sludge cake after process by CAPB+STAD+CaO was 270 much lower than that after CAPB+STAD process. 271 Thus, CAPB+STAD+CaO 272 process could simultaneously enhance biopolymers production and sludge dewaterability. 273

274 3.4 Adsorption capacity of the produced the biopolymers for Cu^{2+}

Fig.5 shows the adsorption capacity of Cu^{2+} by the biopolymers extracted from the three methods and the Pearson correlation between the adsorption capacity of Cu^{2+} and the proportion of PN, PS, and NA in these biopolymers. The biopolymers extracted after CAPB+STAD+CaO process showed the maximum adsorption capacity of 371-392 mg/g for Cu^{2+} , followed were treated by CAPB+STAD and STAD+CaO

with the adsorption capacity of 319-370 mg Cu^{2+}/g biopolymers and 303-333 mg 280 Cu^{2+}/g biopolymers, respectively. Linear fitting results showed that the adsorption 281 capacity of Cu²⁺ has a strong and nearly linear relationship ($R^2 = 0.956$, P < 0.01) with 282 PN proportion in the biopolymers, but the linear regression between the adsorption 283 capacity of Cu²⁺ and PS ($R^2 = 0.216$, P = 0.19) and NA ($R^2 = 0.348$, P = 0.14) in the 284 biopolymers are not significant. Results showed that PN are the key fractions in the 285 biopolymers that responsible for the adsorption of Cu^{2+} , which is also consistent with 286 previous study (Zhou et al., 2016c). CAPB+STAD+CaO process could produce 287 high PN content of the biopolymers, which exhibit the excellent adsorbability for 288 Cu^{2+} . 289

290 *3.5 Mechanisms for simultaneously enhanced biopolymers production and sludge*

291 *dewaterability of WAS by the synergetic integration process*

Fig.6 shows the SEM images of WAS after processed by the three methods, and 292 Fig.7 presents the effect mechanisms of CAPB+STAD, STAD+CaO and 293 CAPB+STAD+CaO on biopolymer productions and sludge dewaterability. Previous 294 study (Zhang et al., 2016) demonstrated that STAD could enhance the production of 295 the biopolymers from WAS. CAPB contains a long alkyl chain, which can form 296 micelles that are effective for improving the solubility of these insoluble 297 macromolecular organic matters in EPS and even IPS of WAS (Fig.6), which is 298 consistent with previous study (Sengco et al., 2001). Thus, CAPB+STAD process 299 could dramatically enhance the production of the biopolymers and its fractions (Fig.1 300 301 (a)-(d)). However, the normalized CST of WAS and water content of sludge cake increased during this process, led to a deteriorated dewatering performance and 302

303 content. For one thing, large amounts of solubilized hydrophilic organic matters in supernatant could adsorb free water and then cause poor dewaterability of WAS 304 (Chen et al., 2001). Additionally, due to the high compressibility and low strength, the 305 permeable filter layer of WAS dramatically decreased and even disappeared during 306 the high pressure extrusion process (Thapa et al., 2009; Chen et al., 2010), which 307 prevent the solid-liquid separation and lead to the deteriorated dewaterability of WAS. 308 CaO could increase the solution pH, which could enhance solubilization of 309 310 organic matters and lead to the increase of the concentrations of the biopolymers and its fractions (Liu et al., 2012), but which were markedly lower than that after 311 Moreover, CaO could form a multi-grid processed by CAPB+STAD process. 312 skeleton by interact with the biopolymers in WAS, which enhances the strength and 313 314 reduces the compressibility of WAS, and the permeable filter layer of WAS still exist during the high pressure extrusion process (Thapa et al., 2009). Thus, CaO could 315 dramatically enhance the dewaterability of WAS (Fig.3 (c)-(d)). 316

317 In the CAPB+STAD+CaO process, CAPB could significantly increase the 318 solubility of macromolecular organic matters in WAS and then dramatically enhances 319 the biopolymers production. With subsequent addition of 0.1~0.2 g CaO/g TSS to the digested sludge, both of the biopolymers and PN concentrations almost remained 320 stable at about 52 and 32 mg/g TSS, respectively, and the biopolymers showed the 321 adsorption capacity of about 375 mg/g for Cu^{2+} . Additionally, CaO could form a 322 323 multi-grid skeleton and dramatically enhance the dewaterability of WAS. For 324 example, the water content of sludge cake dramatically decreased from 78.3% to 74.9% 325 after adding 0.1 g/g TSS of CaO. Thus, CAPB+STAD+CaO process could

326 cost-effectively enhance both the biopolymers production and the sludge327 dewaterability.

For the sustainable development of both environment and society, the 328 indiscriminate discharge of surfactant-containing wastewater after WAS reduction is 329 not acceptable due to its toxicity to human beings and the environment. 330 Thus, CAPB as a biodegradable amphoteric surfactant was selected. It has been 331 demonstrated in the previous work that 91.2% of CAPB with the initial concentration 332 of 0.08 g/g dry sludge could be biodegraded after 24 h by STAD system (Zhou et al., 333 2017d). Moreover, another previous study has also demonstrated continuous 334 biodegradation of a set of surfactants using an oxygen-based membrane biofilm 335 reactor (O₂-MBfR). Bubbleless O₂ transfer completely eliminated foaming, and 336 biofilm accumulation helped the surfactant biodegraders resist toxicity (Lai et al., 337 2017). Thus, the surfactant biodegradation should prevent their discharge to the 338 aquatic environment. 339

340

341 4 Conclusions

Investigating the effect of CAPB+STAD+CaO process on biopolymers production and sludge dewaterability, we found that CAPB could significantly increase the solubility of macromolecular organic matters in WAS and then dramatically enhance the biopolymers. CaO (0.1~0.2 g/g TSS) could dramatically enhance sludge dewaterability by forming a multi-grid skeleton in WAS, while the biopolymers production could almost remain stable at 52 mg/g TSS. The biopolymers showed strong adsorbability for Cu²⁺ (375 mg/g biopolymers, and PN

349	was the key fractions responsible for the biosorption. Especially, the synergetic
350	integration process of STAD with 0.1 g CAPB/g TSS and for 8 h and 0.1 g CaO/g
351	TSS could cost-effectively enhance both the biopolymers production and the sludge
352	dewaterability.
353	
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364

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365 Abbreviations

361

362

- 366 WAS, waste activated sludge;
- 367 CAPB, cocoamidopropyl betaine;
- 368 STAD, short-time aerobic digestion;
- 369 CaO, calcium oxide;
- 370 WWTPs, wastewater treatment plants;
- 371 EPS, extracellular polymeric substances;

- 372 PN, proteins;
- 373 PS, polysaccharides;
- 374 NA, nucleic acids;
- 375 TSS, total suspend solids;
- 376 VSS, volatile suspension solid;
- 377 DO, dissolved oxygen;
- 378 EPS, extracellular polymeric substances;
- 379 IPS, intracellular polymeric substances;
- 380 SEM, scanning electron microscopy.

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496 TABLES

activated studge used in this study							
Unit	Value						
-	6.9 ± 0.2						
g/L	8.04 ± 0.28						
g/L	5.48 ± 0.31						
s·L/g	5.83 ± 0.36						
g/L	9.61 ± 0.12						
mg/L	74.1 ± 11.4						
	Unit - g/L g/L s·L/g g/L						

Table 1 The parameters of concentrated waste activated sludge used in this study a^{a}

^{*a*} TSS, total suspended solids; VSS, volatile suspended solids; CST, capillary suction time normalized by sludge concentration; TCOD, total chemical oxidation demand; SCOD, soluble chemical oxidation demand.

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Table 2 Parameters of the linear relationship between the CAPB dosage and the concentrations of the biopolymers and its fractions at the noted aerobic digestion times ^{*a*}

т.	Proteins		Polysaccharides			Nucleic acids		Biopolymer	
Time (h)	K (mg/g)	R^2	K (mg/g)	R^2	_	K (mg/g)	R^2	K (mg/g)	R^2
0	171	0.970	76.2	0.968		113	0.960	360	0.968
2	201	0.974	80.0	0.988		142	0.955	424	0.973
4	231	0.969	91.4	0.981		163	0.957	486	0.973
8	238	0.958	108	0.979		173	0.956	520	0.969
12	230	0.954	99.2	0.975		175	0.951	511	0.961
24	214	0.955	93.0	0.970		151	0.952	486	0.963

^{*a*} K is the slope of the linear relationship between the CAPB dosage and the concentrations of biopolymer and its fractions. y = Kx+m, where y is concentrations of the biopolymers and its fractions, x is the dosage of CAPB, and m is the intercept.

504 FIGURES



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Figure 1 Effects of CAPB dosage on the production of (a) proteins, (b)
polysaccharides, (c) nucleic acids, and (d) biopolymers and the (e) normalized CST of
WAS and (f) water content of sludge cake at the noted aerobic digestion times.



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Figure 2 Pearson correlations between the normalized CST and the concentrations of

513 proteins, polysaccharides, nucleic acids and the biopolymers in the filtrates.



Figure 3 Effects of CaO dosage on the (a) production of biopolymers and its fractions,
(b) fractions proportion in biopolymers, (c) normalized CST of WAS and (d) water
content of sludge cake at aerobic digestion time of 8 h. PN, PS and NA represent
proteins, polysaccharides and nucleic acids, respectively.



Figure 4 Synergetic effects of the STAD with CAPB and CaO on the (a) production of 522 the biopolymers and its fractions, (b) fractions proportion in biopolymers, (c) 523 524 normalized CST of WAS and (d) water content of sludge cake under the CAPB 525 dosage of 0.10 g/g TSS and aerobic digestion time of 8 h. PN, PS and NA represent 526 proteins, polysaccharides and nucleic acids, respectively.





Figure 5 (a) Adsorption capacity of Cu^{2+} by the biopolymers extracted from the three processes and the Pearson correlation between the adsorption capacity of Cu^{2+} and the proportion of (b) proteins, (c) polysaccharides, and (d) nucleic acids in biopolymers.



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Figure 6 SEM images of waste activated sludge at the aerobic digestion of 8 h: (a) control, (b) adding 10% of CAPB, adding 10% of CAPB and 30% of CaO with the scale of (c) 80 µm and (d) 1µm

scale of (c) 80 μ m and (d) 1 μ m.



- 537 Figure 7 Mechanisms of simultaneously enhanced biopolymer productions and sludge
- 538 dewaterability for WAS by the synergetic integration process of STAD with CAPB
- and CaO.

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Highlights

- Simultaneous biopolymers production and sludge dewaterability of WAS was studied
- Short-time aerobic digestion (STAD) improved biopolymers production by biosynthesis
- CAPB further promoted STAD of WAS to enhance biopolymers production
- CaO enhanced sludge dewaterability but had minor effect on biopolymers

production

• STAD with CAPB and CaO enhanced both biopolymers production and WAS

dewaterability